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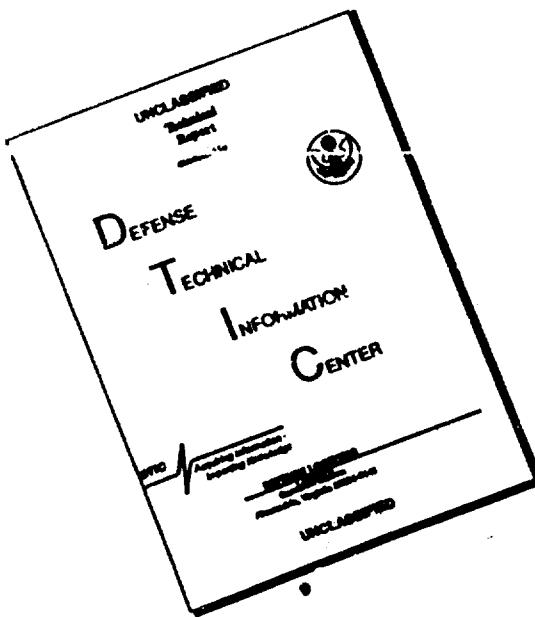
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On the chemical composition of the antigenic substances of the agent of tularemia.

In this report the virulent strain, No. 21, of the bacterium *Tularemia*, representing the S-form of microbe, was utilized. The cultures were cultivated for 3 days at 37°C. on a fish agar with cystine and glucose. The antigenic substances were extracted from a rinsed, fatless, dry micrelial mass.

The antigenic substances of *B. tularemiae* were extracted by the following methods: I--extraction with 5% trichloracetic acid at 0°C. for 3 hours (Duaven method)(1); II--extraction by this same acid but for longer periods of time and at a higher temperature (modified Duaven method); III--extraction by phenol (A. P. Monikov method)(1). The average outcome of the antigenic substances were: for I—3.84%; for II—5.95%; for III—3.24%. The extracted substances were rinsed many times by reprecipitation of them with alcohol from water solutions. Subsidiary workin^gs of the obtained preparations with chloroform (2) indicated an absence of any mechanical tinge of albumin in them. The extracted substance possessed antigenic and allergenic properties, and also a serological activity, which gave a precipitating reaction in great dilutions on anti-tularemia serums.

An attempt was made to decompose the antigens into separate components by means of heating for 1 hour in a boiling water bath, in the presence of 0.1L of acetic acid. Unfortunately the attempt was without results. Exchanging the hydrolysates with heated chloroform, we were able to extract an abundant residue from them, which dissolved well in

a mixture of equal parts of heated chloroform and ethyl alcohol. After removal of the chloroform extracted residue, the hydrolysate was worked with various quantities of ethyl alcohol.

As a result of the above working the antigens of the tularemia microbe were divided into 5 fractions: I-substances non-soluble in a mixture of alcohol-chloroform (6.0-7.3%); II-residue from 4 volumes of alcohol (23.3-35.2%); III-residue from 10 volumes of alcohol (15.4-15%); IV-substances extractable from the hydrolysate by chloroform and ether, also substances which entered into a mixture of alcohol-chloroform during the obtaining of fraction I (33.9-37.8%); V- dry balance after removal of all the above fractions (11.8-18.2%). The antigens, extracted by various conditions of trichloroacetic acid, were characterized by a similar content of separate fractions. The antigen obtained by the Konikova method differed from them somewhat.

A chemical analysis of the original antigens and products of their fractionation indicated that during hydrolysis with acetic acid the separation of the antigenic substances into chemically individual components (specific polysaccharides and albumin) does not take place (Table 1). Each fraction, except the lipid (fraction IV), evidently, represents an intricate mixture of various elements of the antigen, or complexes containing carbohydrate as well as nitrous elements. Nucleic acids were detected in the composition of fraction II.

The studying of the quantitative composition of the carbohydrate and albuminous components of the original antigen, and their separate fractions, was done by chromatography on paper.

The composition of the monosaccharides, obtain & as a result of hydrolysis, was studied on single dimensional chromatograms, taking place in a mixture of n-butanol-acetic acid-water(40:10:50). As a

developer we used anilinphthalate in water-impregnated butanol. For the identification of the glucose and galactose we, besides this, used reactions with phenylhydrazine, having in mind that the osazones of these monose are slightly different.

It was proven that all the antigens, irregardless of the method used for their extraction, contain quantitatively similar specific polysaccharides. Results of the chromatographic analysis of the polysaccharides of the antigens isolated by the Buaven method, and the fractions II and III obtained from them, indicate that the carbohydrate components of the whole antigen and its fraction II are qualitatively of the same composition. They contain the following monosaccharides: galactose, arabinose, xylose, uronic acids and hexosamine. In the composition of the carbohydrate component of fraction III there were other saccharides detected, and namely: glucose, mannose, xylose and hexosamine (analytically).

It is necessary to note that chromatographic analysis of the composition of the polysaccharides of the original antigen disclosed no glucose or mannose. Judging by the intensiveness of the color of the stains on the chromatograms, the basic mass of the polysaccharides of the original antigen consists of galactose and arabinose. The intensively dyeing stains of these sugars on the chromatograms could have masked the weakly dyeing stains of the mannose and glucose.

Chromatographic study of the albuminous components of the antigenic complexes, extracted by various methods from a virulent strain of *B. tularensis*, indicated that they have a qualitatively similar amino-acid content. Separation of the aminoacids of the hydrolysates being studied was done on one- and two-dimensional chromatograms. Water impregnated butanol was used in a mixture of n-butanol-acetic acid-water

(40:10:50) as a solvent during the two-dimensional chromatography. An 0.2% solution of ninhydrin was used in water-saturated butanole as developer. The aminoacidic composition of the albumin of the antigen, extracted by the Buaven method, is represented on the two-dimensional chromatogramm. 15 stains were located on the chromatogramm, of which one was unidentified. It is believed that it belongs to α,ϵ -diaminopymeric acid. Histidine and lysine are located on this chromatogramm in one stain(spot). Their presence was indicated on the single-dimensional chromatograms. The histidine, besides this, was detected by the Paul reaction. The presence of proline was indicated on a separate chromatogramm during the use of isatin. Consequently, the albuminous components of the antigen contain more than 17 aminoacids. In regard to the aminoacidic composition of the albuminous component, fraction II and III did not differ from each other.

The conducted studies established that the antigenic complex of the tularemia microbe, by its chemical nature, significantly differs from the full antigens of Buaven, from the bacteria of the intestinal-typhus group, studied to this time. Two complicated complexes-fractions II and III- enter into the composition of the tularemia antigen.

It was established that the latter complexes are separate components, firmly linked together. Thus, fraction II represents a stable complex, consisting of polysaccharides, albumin and nucleic acid, and fraction III-stable polysaccharides, albuminous complex. The albuminous components of the said fractions contain at least 17 similar amino-acids. The presence of qualitatively different polysaccharides in these fractions indicates: either two antigens, or one with various determinant groups.

Great interest is shown in the detection of an exceptionally high content of the lipid fraction in the tularemia antigen(approx. 40% from the original antigen), sharply differing from the analogical fractions of the Buaven full antigens, by characteristics.

INST. EPID. MICROB. imene H. F. GAMALE. AMS., USSR.
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Literature

1. V. I. Govarnitski, J.MI., 2, 37(1947); 10, 31 and 11, 82(1948).
2. M. G. Savage, D. B. Zackman, A. Z. Smolens. J. Biol. Chem., 124, 425, (1938). 3. A. N. Bolezerski, N. I. Proskuryakov, Prac. Manual for the Biochemistry of Plants, 1951. 4. S. M. Partridge, Nature, 164, 443 (49).
5. L. Elson, W. Morgan, Biochem. J., 27, 1824(1933).

Table 1. Chemical characteristics of antigens and their separate fractions (data expressed in μ fraction weight of prep.)

Chemical indicators	Orig. antigen		I fraction		II fraction		III fraction		IV fraction		V fraction	
	B	K	D	H3	K	B	hB	K	B	hB	K	B
Urea nitrogen	7.1	7.4	5.9	8.6	8.5	5.7	8.7	8.5	9.5	9.5	16.1	2.5
Urea phospho- rus	1.0	1.2	0.9	0.8	1.4	0.6	1.7	1.5	1.9	0.6	0.6	1.2
Phosphorus nucleic acid **								1.5	1.5			
Oxyribonucleic acid ***								5.2	5.1			
Nucleic acid ****								10.5	10.9			
using sub- stances	20.7	20.2	20.1	7.1	5.1	6.9	27.9	28.5	26.7	25.5	25.0	25.4
Urea	11.0	10.9	9.6	0	0	0	16.8	13.4	14.0	20.3	20.2	19.6
Urea hexose (by calcu- lation)	0.9	0.9	0.8	0	0	1.3	1.1	1.1	1.6	1.6	1.5	0.2
Urea (by cal- culation)						1.7	1.8					
Antigen during calculation												
Antigen (by cal- culation)												

* Antigens obtained: H-juven, H-juven modified Juven method, K-Korlikov.

** Data of the substance were determined only in fraction II.

*** Fractionation of the phosphorus combination according to Shmidt and Tanyuk (3).

**** Fractionation of the phosphorus combination according